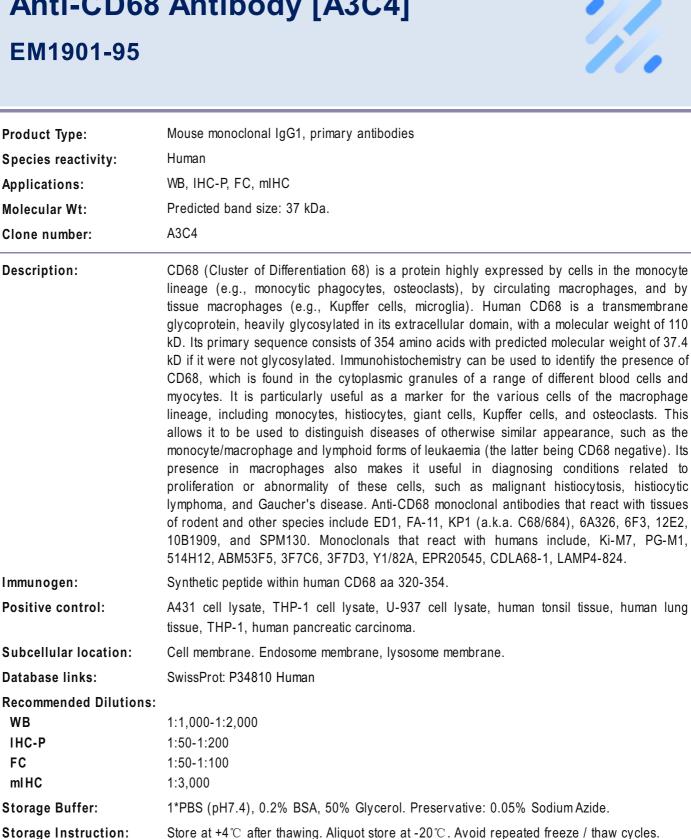
Anti-CD68 Antibody [A3C4]



Protein G affinity purified. Purity:

Hangzhou Huaan Biotechnology Co., Ltd.

Orders:0086-571-88062880

Technical:0086-571-89986345

Service mail:support@huabio.cn



Applications:WB=Western blot IHC-P=Immunohistochemistry (paraffin) IF-Cell=Immunofluorescence (Cell) IF-Tissue=Immunofluorescence (Tissue) FC=Flow cytometry IP=Immunoprecipitation

Images

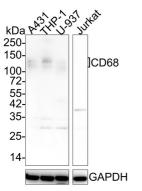


Fig1: Western blot analysis of CD68 on different lysates with Mouse anti-CD68 antibody (EM1901-95) at 1/1,000 dilution.

Lane 1: A431 cell lysate Lane 2: THP-1 cell lysate Lane 3: U-937 cell lysate Lane 4: Jurkat cell lysate (negative)

Lysates/proteins at 20 µg/Lane.

Predicted band size: 37 kDa Observed band size: 100-150 kDa

Exposure time: 3 minutes;

4-20% SDS-PAGE gel.

Proteins were transferred to a PVDF membrane and blocked with 5% NFDM/TBST for 1 hour at room temperature. The primary antibody (EM1901-95) at 1/1,000 dilution was used in 5% NFDM/TBST at 4° C overnight. Goat Anti-Mouse IgG - HRP Secondary Antibody (HA1006) at 1/50,000 dilution was used for 1 hour at room temperature.

Fig2: Immunohistochemical analysis of paraffin-embedded human tonsil tissue with Rabbit anti-CD68 antibody (EM1901-95) at 1/1,000 dilution.

The section was pre-treated using heat mediated antigen retrieval with Tris-EDTA buffer (pH 9.0) for 20 minutes. The tissues were blocked in 1% BSA for 20 minutes at room temperature, washed with ddH₂O and PBS, and then probed with the primary antibody (EM1901-95) at 1/1,000 dilution for 1 hour at room temperature. The detection was performed using an HRP conjugated compact polymer system. DAB was used as the chromogen. Tissues were counterstained with hematoxylin and mounted with DPX.

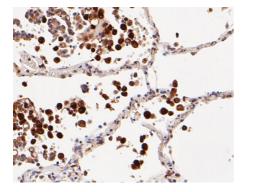


Fig3: Immunohistochemical analysis of paraffin-embedded human lung tissue using anti-CD68 antibody. The section was pre-treated using heat mediated antigen retrieval with Tris-EDTA buffer (pH 8.0-8.4) for 20 minutes. The tissues were blocked in 5% BSA for 30 minutes at room temperature, washed with ddH₂O and PBS, and then probed with the primary antibody (EM1901-95, 1/200) for 30 minutes at room temperature. The detection was performed using an HRP conjugated compact polymer system. DAB was used as the chromogen. Tissues were counterstained with hematoxylin and mounted with DPX.

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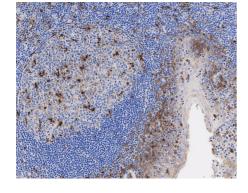


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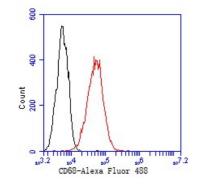


Fig4: Flow cytometric analysis of CD68 was done on THP-1 cells. The cells were fixed, permeabilized and stained with the primary antibody (EM1901-95, 1/50) (red). After incubation of the primary antibody at room temperature for an hour, the cells were stained with a Alexa Fluor 488-conjugated Goat anti-Mouse IgG Secondary antibody at 1/1000 dilution for 30 minutes.Unlabelled sample was used as a control (cells without incubation with primary antibody; black).

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Fig5: Fluorescence multiplex immunohistochemical analysis of the pancreatic carcinoma (Formalin/PFA-fixed paraffinhuman embedded sections). Panel A: the merged image of anti-CD68 (EM1901-95, green), anti-CD163 (ET1704-43, red) and anti-PanCK (HA601094, violet) on human pancreatic carcinoma. Panel B: anti- CD68 stained on M1 macrophages. Panel C: anti-CD163 stained on M2 macrophages cells. Panel D: anti-panCK stained on cancer cells. HRP Conjugated UltraPolymer Goat Polyclonal Antibody HA1119/HA1120 was used as a secondary antibody. The immunostaining was performed with the Sequential Immunostaining Kit (IRISKit[™]MH010101, www.luminiris.cn). The section was incubated in three rounds of staining: in the order of EM1901-95 (1/3,000 dilution), ET1704-43 (1/3,000 dilution), and HA601094 (1/3,000 dilution) for 20 mins at room temperature. Each round was followed by a separate fluorescent tyramide signal amplification system. Heat mediated antigen retrieval with Tris-EDTA buffer (pH 9.0) for 30 mins at 95℃. DAPI (blue) was used as a nuclear counter stain. Image acquisition was performed with Nikon ECLIPSE Ni-E microscope.

Note: All products are "FOR RESEARCH USE ONLY AND ARE NOT INTENDED FOR DIAGNOSTIC OR THERAPEUTIC USE".

Background References

- 1. Wang L. et. al. Specific clinical and immune features of CD68 in glioma via 1,024 samples. Cancer Manag Res. 2018 Nov 27;10:6409-6419.
- 2. Minami K. et. al. Prognostic significance of CD68, CD163 and Folate receptor-β positive macrophages in hepatocellular carcinoma. Exp Ther Med. 2018 May;15(5):4465-4476.

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